

# Communication

Subscriber access provided by The Libraries of the | University of North Dakota

# Spirodiepoxides in Total Synthesis: Epoxomicin

Sreenivas Katukojvala, Kristin N. Barlett, Stephen D. Lotesta, and Lawrence J. Williams J. Am. Chem. Soc., 2004, 126 (47), 15348-15349• DOI: 10.1021/ja044563c • Publication Date (Web): 04 November 2004 Downloaded from http://pubs.acs.org on April 5, 2009



## **More About This Article**

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Links to the 3 articles that cite this article, as of the time of this article download
- Access to high resolution figures
- Links to articles and content related to this article
- · Copyright permission to reproduce figures and/or text from this article

View the Full Text HTML





Published on Web 11/04/2004

## Spirodiepoxides in Total Synthesis: Epoxomicin

Sreenivas Katukojvala, Kristin N. Barlett, Stephen D. Lotesta, and Lawrence J. Williams\*

Department of Chemistry and Chemical Biology, Rutgers, The State University of New Jersey, Piscataway, New Jersey 08854

Received September 8, 2004; E-mail: ljw@rutchem.rutgers.edu

Substrate-directed and reagent-controlled asymmetric alkene epoxidation and subsequent nucleophilic epoxide opening are among the most widely used strategies for introducing molecular complexity in target-oriented synthesis.<sup>1</sup> Yet the analogous oxidation/nucleophilic opening of the intrinsically stereogenic allene remains unexplored in synthesis.<sup>2</sup> The oxidation products, spirodiepoxides (e.g., **3**), can in principle serve as synthetically useful three-carbon units of bond formation and stereochemistry and provide direct access to highly functionalized and highly enantioenriched ketones and ketone derivatives. Here we present an efficient route to the potent and selective proteasome inhibitor epoxomicin<sup>3</sup> (**1**) and thus establish the first use of the spirodiepoxide functional group in total synthesis.

Proteasome targeting has emerged as a new modality for the potential treatment of diseases ranging from malaria to cancer.<sup>4</sup> The importance of understanding and controlling proteasome function led us to design new approaches to epoxomicin (1).<sup>5</sup> In the most concise approach, the functionality and stereochemical arrangement of simplified structure 2, present in 1, would be orchestrated simultaneously (Scheme 1), and modifications in the peripheral highlighted portions of 2 would not encumber the synthetic route. Spirodiepoxides of type 3 could serve as precursors to such target systems. In the presence of a suitable nucleophile such species should undergo regioselective S<sub>N</sub>2 reactions. Enantiomerically pure spirodiepoxide would be derived from oxidation of optically pure, and appropriately protected, allene (e.g., 5), which would arise in turn from aldehyde, alkyne, and organometallic precursors. The highlighted substituents of these precursors correspond to those indicated in 2. Oxidation of 5, first at the less hindered face of the more substituted  $\pi$ -bond, would give rise to 3 as the major isomer. This approach would require identification of a hydroxyl protecting group for 5 and a nitrogen nucleophile suitable for spirodiepoxide opening.

The pioneering work of Crandall stands as the only reported systematic analysis of spirodiepoxides.<sup>6</sup> Thus, simple allenes may be oxidized to spirodiepoxides, which rearrange in the presence of acid. A number of nucleophiles have been successfully added to spirodiepoxides, and consistent with  $S_N 2$  substitution, the ratio of products corresponds to the ratio of oxidation products.

To initiate our studies, a series of *O*-protected hydroxy allenes (6-13) was prepared and oxidized with dimethyldioxirane (DMDO), as shown in Scheme 2. Functional group compatibility and approximate ratios of oxidation products were determined by <sup>1</sup>H NMR analysis of the crude spirodiepoxides.<sup>7</sup> While benzoate 13 led to ortho ester 14, silyl and alkyl ether protection is compatible with the spirodiepoxide functionality and gave the corresponding spirodiepoxides in ratios of approximately 2:1.

To proceed toward **1**, suitable nitrogen nucleophiles were identified by exposure to spirodiepoxides derived from allene **6**.<sup>7</sup> Addition of benzamide under neutral conditions led to *O*-alkylation/ cyclization and gave oxazolines **15** in 44% yield, whereas the

## Scheme 1





<sup>*a*</sup> Reagents and conditions: (i) DMDO, acetone, -40 to 23 °C, 2 h; then nucleophile (see Supporting Information); (ii) DMDO, acetone, -50 to 23 °C, 2 h; (iii) Bu<sub>4</sub>NN<sub>3</sub>, CHCl<sub>3</sub>, -30 °C, 2 h; 30%. Major isomers shown. SES = SO<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>Si(CH<sub>3</sub>)<sub>3</sub>.

*N*-alkylated product **16** predominated, albeit in modest yield (30%), when benzamide was first treated with *n*-BuLi. Benzenesulfonamide in the presence of base<sup>8</sup> gave adduct **17** (75%), but under neutral conditions no product was observed. In hopes of inducing *N*-alkylation of an amide precursor, we prepared *N*-acyl sulfonamide **18** by using our thio acid/azide amidation.<sup>9</sup> Under a variety of conditions (DIEA, K<sub>2</sub>CO<sub>3</sub>, LiHMDS, NaHMDS, or KHMDS), no reaction with the spirodiepoxide took place. In the absence of base, however, an unstable adduct formed and then decomposed upon treatment with fluoride. While sodium azide added slowly to spirodiepoxides derived from **6**, tetrabutylammonium azide added rapidly even at low temperature to give **19** in 73% yield.

When the spirodiepoxide derived from allenyl mesylate **20** was exposed to tetrabutylammonium azide, epoxide **22** was formed in 30% yield. Azide appears to preferentially open the spirodiepoxide rather than displace the neopentyl-like mesylate (see **21**). The nascent vicinal alkoxide displaces the mesylate to form a new epoxide, which resists subsequent azide opening. Azide **22** and the amine derived by reduction proved unstable and therefore were not

#### Scheme 3<sup>a</sup>



<sup>a</sup> Reagents and conditions: (i) (-)-N-methylephedrine, Zn(OTf)<sub>2</sub>, Et<sub>3</sub>N, toluene, rt, 2 h, TBSOCH<sub>2</sub>CCH then 23 14 h, 93%, >95% ee; (ii) a. MsCl, Et<sub>3</sub>N, DCM, -65 to 23 °C, 2 h; b. MeMgBr, CuBr, LiBr, THF/tert-butyl ether, -65 to 23 °C, 2 h, 91%; (iii) a. DMDO, -40 to 23 °C, 1.5 h; b. Bu4NN3, CHCl3, -20 to 23 °C, 1 h, 73% (3:1 dr); c. 10% Pd/C, H2, (Boc)<sub>2</sub>O·K<sub>2</sub>CO<sub>3</sub>, EtOAc, rt, 12 h, 91%; d. TFA, 0 °C, 13 min; (iv) a. 26, HCl·Ile-OMe, DCC, HOBT, Et<sub>3</sub>N, DMF, 0 to 23 °C, 12 h, 93%; b. 25% TFA-DCM, 10 to 23 °C, 40 min; c. TEA, Ac<sub>2</sub>O, DMAP, DCM, 0 to 23 °C, 3 h, 95%; d. 5% NaOH, MeOH-H2O, rt, 2 h, 99%; e. 27, DCC, HOBT, DCM-DMF, rt, 3 h, 92%; f. 10% Pd/C, H<sub>2</sub>, MeOH, rt, 2 h, 100%; (v) 25, DIEA, DCC, HOBT, DCM-DMF, rt, 4 h, 86%; (vi) a. TBAF, THF, 0 to 23 °C, 1 h, 89%; b. MsCl, DIEA, DCM, -40 to 23 °C, 1 h; c. K<sub>2</sub>CO<sub>3</sub>, THF-H<sub>2</sub>O, rt, 3 h, 93%; d. TFA, 0 to 23 °C, 20 min, 88%.

advanced. Amines related to 19, however, were stable to manipulation leading to 1, as described below.

The optimized synthesis of epoxomicin is presented in Scheme 3. Isovaleraldehyde was subjected to asymmetric alkynylation<sup>10</sup> to form 24 (93% yield, >95% ee). The alcohol was converted to the mesylate and subsequently transformed into allene 6 upon coppermediated<sup>11</sup> S<sub>N</sub>2' displacement (91%). As described above, treatment of 6 with DMDO<sup>12</sup> followed by exposure to azide smoothly produced 19 (>95% er) in 73% yield (3:1 ratio of separable diastereomers). In situ reduction/protection (91%) and then treatment with acid converted the major azido alcohol to the stable crude amine salt (25) ready for peptide coupling. DCC-promoted coupling of 26 with methyl isoleucinate (93%), Boc removal and acetylation (95%), saponification (99%), coupling to threonine 27<sup>7</sup> (see inset), and then hydrogenolysis gave 28 (92%, two steps), which smoothly coupled with 25 to furnish 29 (86%,  $19 \rightarrow 29$ ). Exposure of 29 to fluoride cleaved the silvl ether-protecting group (89%). The resultant primary alcohol was converted to the mesylate and cyclized to give the epoxide (93%), and then the *tert*-butyl ether was removed (88%) to produce 1.

Spectral data for synthetic 1 proved identical to published data for natural 1, including  $[\alpha]_D^{25}$  -64.0 (c 0.47, MeOH), lit<sup>3a</sup> -66.1 (c 0.50, MeOH), and confirms the stereochemical assignments in Scheme 3. Chemical correlation secured the syn stereochemistry of the major isomers of 15-17, 19, and 22 as shown in Scheme 2.

Nucleophilic opening of a spirodiepoxide effectively establishes specific stereochemical communication across a carbonyl. As depicted in eq 1, oxidation/nucleophilic opening installs three



functional groups, nucleophile, ketone, and alcohol, with syn

selectivity. Importantly, this transformation is achieved in the absence of other stereodirecting functionalities. This report also establishes that chiral spirodiepoxides can be prepared and manipulated on gram scale.13 Coupled with the two-step assembly of all the carbon atoms of the targeted substructure, this method is a highly efficient, flexible, and modular synthesis of highly functionalized ketones and their derivatives. In addition to disclosing the selective reaction of a spirodiepoxide in the presence of a mesylate and several new and diverse reactions of spirodiepoxides (e.g. formation of ortho ester 14, oxazoline 15, and azido epoxide 22), we have completed the synthesis of epoxomicin in an overall yield of 26% from 23 (20% including all steps). This route should also provide new epoxomicin analogues with improved activity and selectivity. Issues such as the scope of compatible functional groups and nucleophiles, control of regioselective nucleophilic addition, stereoselective oxidation of the allene precursors, and elucidation and exploitation of the mechanisms of spirodiepoxide opening are under investigation.

Acknowledgment. Financial support by Merck & Co., Johnson & Johnson (Discovery Award), donors of the American Chemical Society Petroleum Research Fund, Graduate Assistance in Areas of National Need (fellowship to S.D.L.), and Rutgers, The State University of New Jersey is gratefully acknowledged. We thank Prof. Spencer Knapp for stimulating discussions and critical reading of the manuscript.

Supporting Information Available: Synthetic methods and characterization data. This material is available free of charge via the Internet at http://pubs.acs.org.

### References

- (1) Kolb, H. C.; Finn, M. G.; Sharpless, K. B. Angew. Chem., Int. Ed. 2001, 40, 2004–2021. Rao, A. S. In Comprehensive Organic Synthesis; Trost, B. M., Flemming, I., Eds.; Pergamon: Oxford, 1991; Vol. 7, pp 357-387. Hickey, M.; Goeddel, D.; Crane, Z.; Shi, Y. Proc. Natl. Acad. Sci. U.S.A. 2004, 101, 5794–5798.
- (2) For recent related allene oxidations with synthesis applications, see: Xiong, H.; Huang, J.; Ghosh, S. K.; Hsung, R. P. J. Am. Chem. Soc. 2003, 125, 12694–12695. Fleming, S. A.; Carroll, S. M.; Sean, M.; Hirschi, J.; Liu, R.; Pace, J. L.; Redd, J. T. *Tetrahedron Lett.* **2004**, *45*, 3341–3343.
- (3) Hanada, H.; Sugawara, K.; Kaneta, K.; Toda, S.; Nishiyama, Y.; Tomita, K.; Yamamoto, H.; Konishi, M.; Oki, T. J. Antibiot. 1992, 45, 1746– 1752. Meng, L.; Mohan, R.; Kwok, B. H. B.; Elofsson, M.; Sin, N.; Crews, C. M. Proc. Natl. Acad. Sci. U.S.A., 1999, 96, 10403–10408.
- (4) For proteasome structure, biology and applications lead references, see: Kloetzel, P. M. Nat. Rev. Mol. Cell. Biol. 2001, 2, 179-188
- (5) Sin, N.; Kim, K. B.; Elofsson, M.; Meng, L.; Auth, H.; Kwok, B. H. B.; Crews, C. M. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 2283–2288.
  (6) Crandall, J. K.; Machleder, W. H. *Tetrahedron Lett.* **1966**, *7*, 6037–6041. Crandall, J. K.; Batal, D. J. *Tetrahedron Lett.* **1988**, *29*, 4791–4794. Crandall, J. K.; Reix, T. *Tetrahedron Lett.* **1994**, *35*, 2513–2516. Crandall, J. K.; Reix, T. *Tetrahedron Lett.* **1994**, *35*, 2513–2516. Crandall, J. K.; Reix, T. *Tetrahedron Lett.* **1994**, *35*, 2513–2516. J. K.; Rambo, E. Tetrahedron Lett. 1994, 35, 1489-1492. Crandall, J K.; Machleder, W. H. J. Am. Chem. Soc. 1968, 90, 7292-7296. Crandall, J. K.; Machleder, W. H.; Thomas, M. J. J. Am. Chem. Soc. 1968, 90, 7346–7347. Crandall, J. K.; Batal, D. J.; Lin, F.; Reix, T.; Nadol, G. S.; Ng, R. A. *Tetrahedron* **1992**, *48*, 1427–1448. Crandall, J. K.; Rambo, E. Tetrahedron 2002, 58, 7027-7036. Crandall, J. K.; Conover, W. Komin, J. B.; Machleder, W. H. J. Org. Chem. **1974**, *39*, 1723–1729.
   Crandall, J. K.; Batal, D. J. J. Org. Chem. **1988**, *53*, 1338–1340. Crandall, J. K.; Rambo, E. J. Org. Chem. **1990**, *55*, 5929–5930. Crandall, J. K.; Batal, D. J.; Sebesta, D. P.; Lin, F. J. Org. Chem. **1991**, *56*, 1153–1166. (7) See also: Supporting Information.
- Albanese, D.; Landini, D.; Penso, M. Tetrahedron 1997, 53, 4787-4790. Shangguan, N.; Katukojvala, S.; Greenberg, R.; Williams, L. J. J. Am. (9)Chem. Soc. 2003, 125, 7754-7755
- (10) Frantz, D.; Fassler, R.; Carreira, E. M. J. Am. Chem. Soc. 2000, 122, 1806 - 1807
- (11) Elsevier, C. J.; Stehouwer, P. M.; Westmijze, H.; Vermeer, P. J. Org. Chem. 1983, 48, 1103-1105. Elsevier, C. J.; Vermeer, P. J. Org. Chem. 1984, 49, 1649-1650.
- Murray, R. W.; Jeyaraman, R. J. Org. Chem. 1985, 50, 2847-2853.
- See also: Rameshkumar, C.; Xiong, H.; Tracey, M. R.; Berry, C. R.; Yao, L. J.; Hsung, R. P. J. Org. Chem. **2002**, 67, 1339–1345.

JA044563C